Antibody Aliquot Preparation Protocol

A. Preparation of diluted antibodies

1. Spin the antibody vial (500 µl) briefly in the microfuge
2. Add 5 µl 10% NaN₃. Careful, very toxic!
3. Add 500 µl glycerol. Be very careful pipetting, it is very viscous. Mix antibody, glycerol and NaN₃ pipetting up and down gently until you have a perfectly homogeneous solution.
4. Spin the vial briefly. At this point you can store this vial indefinitely at -20°C.

B. Aliquoting

1. Label 50 sterile Eppendorf tubes with “ZzXxx AYYY”, where Z is the species in which the secondary is raised, X is the species against which the antibody is raised and YYY the Alexa dye wavelength. For example, a Goat-anti Rabbit Alexa 568 would become GoRabA568. Donkey anti mouse Alexa 488 becomes DoMouA488.
2. Dispense 20 µl aliquots of the antibody/glycerol/NaN₃ solution into each tube.
3. Place the tubes in a storage box at -20°C labeled “Sample Secondary Antibodies UIC” Add another label listing the antibodies inside the box and the date they were aliquoted.